



Crystal Chem

Mouse IGF-1 ELISA Kit Instructions

For the quantitative determination of IGF-1 in
mouse serum, plasma, and cell culture media

**Catalog #80574
96 Assays**

For research use only. Not for use in diagnostic procedures.

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A. Intended Use

The Mouse IGF-1 ELISA kit is for the quantitative determination of IGF-1 in mouse serum, plasma, and cell culture media. Please read the complete kit insert before performing this assay. The kit is for RESEARCH USE ONLY. It is not intended for use in diagnostic procedures.

B. Introduction

IGF-1 (Insulin-like Growth Factor-1) plays a critical role in regulating the proliferation and differentiation of many cell types. IGF-1's major regulators are growth hormone and nutrition.

C. Principle of the Assay

The Mouse IGF-1 ELISA kit is an ELISA sandwich assay for mouse IGF-1. It utilizes two specific and high affinity antibodies for this protein. IGF-1 is first dissociated from the IGF-BPs via an acidic buffer, After neutralization of the samples, the excess IGF-II occupies the IGF-binding sites of the binding proteins, thus allowing the measurement of resulting free IGF-I. The IGF-1 in the mouse sample binds to the first antibody coated on the microtiter plate. In the following step, the biotinylated and Streptavidin-Peroxidase conjugated second specific anti-IGF-1 antibody binds in turn to the immobilized IGF-1. In the closing substrate reaction, the IGF-1 levels of the samples can be measured by color intensity.

D. Kit Storage

1. Upon receipt of the Mouse IGF-1 ELISA kit, store it at 2-8°C and avoid light exposure (do not freeze the kit or hold it at temperatures above 25°C).
2. The kit should not be used after the expiration date.

E. Assay Materials

E.1. Materials provided

TABLE 1 Contents of the kit

Mark	Description	Amount
MIC	Antibody-coated Microplate (12 x 8)	1 pack
STD1-5	Standards	1 x 5 vials
DIL	Dilution Buffer	1 x 125 mL
CON1-2	Controls	1 x 2 vials
AB CONJ	Antibody Conjugate	1 x 7 mL
ENZ CONJ	Enzyme Conjugate	1 x 12 mL
WASH	Wash Buffer (20X Concentrate)	1 x 50 mL
SUB	Substrate Solution	1 x 12 mL
STOP	Stop Solution	1 x 12 mL
	Sealing tape for plate	2x, adhesive

E.2. Materials required but not provided

Micropipettes and disposable tips
Distilled or deionized water
Polypropylene microtubes
Volumetric flasks
Vortex mixer
Microplate shaker (350 rpm)
Microplate reader (capable of reading A_{450} and A_{630} values)

F. Assay Precautions

1. Only appropriately-trained personnel should use the kit. Laboratory personnel should wear suitable protective clothing. All chemicals and reagents should be considered potentially hazardous. Avoid ingestion and contact with skin and eyes.
2. Some assay components may contain human sourced materials. Accordingly, all assay components should be handled as if potentially infectious using safe laboratory procedures.
3. Do not use the reagents after the expiration date.
4. Reagents are light sensitive and should be protected from sunlight.

G. Maximizing Kit Performance

1. Given the small sample volumes required (5 μL), pipetting should be done as carefully as possible. A high quality 10 μL or better precision pipette should be used for such volumes. Drops of liquid adhering to the outside of the pipette tips should be removed by wiping to ensure the highest degree of accuracy.
2. In order to prevent the microplate wells from drying out and to get the best results, samples and reagents should be dispensed quickly into the wells.
3. Each standard and sample should be assayed in duplicate.
4. The same sequence of pipetting and other operations should be maintained in all procedures.
5. Do not mix reagents that have different lot numbers.

H. Sample Collection

Mouse serum and plasma samples collected with EDTA, Citrate, or Heparin anticoagulant can be used. Cell culture media can also be used as a sample matrix after predilution of 1:2 with Dilution Buffer. Hemolytic reactions should be avoided. The blood has to be allowed to clot and after complete clotting, serum is separated by centrifugation. Samples should be chilled as soon as possible after sample withdrawal. For long-term storage, samples can be stored for more than 2 years at -20°C . Avoid repeated freeze-thaw cycles of samples.

I. Assay Procedure

I.1. Preparation of reagents

1. Antibody-coated microplate
Provided as ready to use. Protect from moisture.
2. Standards 1-5
Standards are provided in lyophilized form with concentrations ranging from 0.5 ng/mL to 18 ng/mL. Dilute each standard with 1 mL of Dilution Buffer (marked "DIL"). After reconstitution, it is recommended that standards be allowed to sit for 15 mins at room temperature and then mixed thoroughly but gently with a Vortex mixer. Reconstituted standards are stable for two months at -20°C . Standards should not be repeatedly thawed (at most, once), so standards should be appropriately aliquoted in appropriate volumes prior to

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being frozen. Standards are provided in the following concentrations: 0.5, 2.5, 6, 12, and 18 ng/mL.

3. Dilution Buffer
Provided as ready to use.
4. Controls 1-2
Controls are provided in lyophilized form with target value and ranges included on their labels. Reconstitute controls with 500 μ L of Dilution Buffer. Reconstituted controls are stable for two months at -20°C .
5. Antibody Conjugate
Provided as ready to use.
6. Enzyme Conjugate
Provided as ready to use.
7. Wash Buffer (20X Concentrated)
The wash buffer has to be diluted 1:20 with distilled or deionized water prior to use. For example, 50 mL of wash buffer must be diluted with 950 mL of distilled or deionized water. Wash buffer is stable for 4 weeks at $2-8^{\circ}\text{C}$ after dilution, so dilute only as needed.
8. Substrate Solution
Provided as ready to use.
9. Stop Solution
Provided as ready to use.

I.2. Dilution of samples and controls

1. Samples and controls need to be diluted with Dilution Buffer for use with the assay. Please note that this section only applies to samples and controls, not standards. Samples should be diluted at least 1:10 in dilution buffer. A sample dilution of 1:100 using 5 μ L of sample is generally suitable. Allow diluted samples to incubate at room temperature for at least 15 minutes. Samples and controls must be used within 2 hours once diluted.

Since mouse IGF-1 levels can vary, dilution ratio may need to be adjusted as appropriate.

I.3. Assay procedure

Prior to running the assay, all reagents should be brought to room temperature for at least 30 minutes. Reagents should be stored at $2-8^{\circ}\text{C}$ immediately after use. Before use, mix the reagents thoroughly by gentle agitation or swirling.

1. Add 50 μ L of Antibody Conjugate in all wells to be used.
2. In each well, add 50 μ L of diluted sample or 50 μ L of standard or 50 μ L of diluted control and mix well by repeated pipetting.
Note: A blank using 50 μ L of Dilution Buffer is recommended.
3. Cover the wells with sealing tape and incubate the plate for 1 hour at room temperature (shake at 350 rpm).
4. Aspirate well contents and wash five times using 300 μ L of Wash Buffer per well. After each wash, remove any remaining solution by inverting and tapping the plate firmly on a clean paper towel.
5. Add 100 μ L of the Enzyme Conjugate in each well.
6. Cover the wells with sealing tape and incubate the plate for 30 minutes at room temperature (shake at 350 rpm).
7. Aspirate well contents and wash five times using 300 μ L of Wash Buffer per well. After each wash, remove any remaining solution by inverting and tapping the plate firmly on a clean paper towel.
8. Add 100 μ L of Substrate Solution in each well.

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9. Incubate the plate for 30 mins in dark room at room temperature.
10. Stop the reaction by adding 100 μ L of Stop Solution.
11. Measure absorbance within 30 minutes using a plate reader (measure A_{450} values and subtract A_{630} values).

I.4. Determining the IGF-1 concentration

1. Using computer software, construct the IGF-1 calibration curve by plotting the mean change in absorbance value for each calibrator (incl. blank) on the Y axis versus the corresponding IGF-1 concentration on the X axis. A higher-grade polynomial, or four parametric logistic (4-PL) curve fit or non-linear regression are suitable for the evaluation.

Note: A calibration curve should be plotted every time the assay is performed.

2. Mouse IGF-1 concentrations in the samples are interpolated using the calibration curve and mean absorbance values for each sample. For diluted samples and controls, the values obtained must be multiplied by the dilution factor to obtain the final IGF-1 concentration. The IGF-1 concentration is expressed in ng/mL.

Note: Samples with high mouse IGF-1 concentrations (ie. fall above the range of the assay) should be further diluted with the dilution buffer and rerun.

J. Performance characteristics

J.1. Assay range

The Mouse IGF-1 ELISA Kit has an assay range from 0.5 – 18 ng/mL. The analytical sensitivity of the assay is 0.029 ng/mL.

J.2. Precision

The assay has a within-run and total precision of CV < 10%.

Warranty

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